

Henry Q. Xiong

## Molecular targeting therapy for pancreatic cancer

Published online: 13 August 2004  
© Springer-Verlag 2004

**Abstract** Pancreatic carcinogenesis is driven by multiple genetic and epigenetic changes. The epidermal growth factor receptor (EGFR) and its downstream signaling pathways, Ras-Raf-MEK-ERK axis, play important roles in pancreatic cancer development. The phosphoinositol 3 kinase (PI3 K)/Akt and the nuclear factor  $\kappa$ B (NF- $\kappa$ B) pathways control both proliferation and resistance to apoptosis of pancreatic cancer. The role of cyclooxygenase (COX) and lipoxygenase (LOX) in the development of pancreatic cancer has been made known recently. The elucidation of these molecular events has led to several distinct therapeutic advances, including therapies that target EGFR, the Ras-Raf-MEK-ERK axis, the COX-2 and LOX pathways, and others. Many novel agents have been developed and are undergoing clinical investigation, such as monoclonal antibodies against EGFR, tyrosine kinase inhibitors (TKIs), farnesyl transferase inhibitors (FTIs), Bay43-9006, CI-1040, CCI-779, celecoxib, and LY293111. This review highlights recent advances in the development of these agents.

**Keywords** Pancreatic cancer · Molecular targeting therapy · EGFR inhibitors · Ras inhibitors · COX-2 inhibitors

---

### Molecular targets in pancreatic cancer

The development, continued growth, and metastasis of pancreatic cancer are driven by multiple genetic and epigenetic changes, including inactivation of tumor suppressor genes and activation of protooncogenes. Some of the important genetic and epigenetic changes that have been targeted for drug development are summarized in Table 1. A key molecular event is the overexpression of the epidermal growth factor receptor (EGFR) and activation of its downstream signaling molecules. EGFR is a receptor tyrosine kinase that undergoes dimerization and activation of its intrinsic kinase upon binding of its ligands. The activated EGFR subsequently recruits and activates its downstream signaling molecules. Ras is a key signaling molecule downstream of EGFR. Ras is activated through a guanine exchange factor, Sos, which is activated through interaction with EGFR and adapters such as Grb 2 and Shc. Subsequently, Ras activates Raf, and a dual specific kinase MEK1, and finally extracellular signal-regulated kinase (ERK), which ultimately translocates to the nucleus and regulates transcription factors. This Ras-Raf-MEK-ERK signaling pathway or axis is conserved among different species, indicating its fundamental role in the normal physiology of cells [5, 65].

Dysregulation of the signal transduction pathways may lead to uncontrolled cell growth and hence tumor development. The role for EGFR and its downstream signaling molecules in tumorigenesis is evidenced by their ability to transform normal cells to a neoplastic phenotype when expressed in mutated, unregulated forms, or when expressed to an abnormally high level. Moreover, activation mutation or overexpression of EGFR and its downstream signaling molecules occurs frequently in a variety of human cancers, including pancreatic cancer. The most recent study indicated that EGFR was detectable in more than 95% of patients with pancreatic cancer [1]. In most cases, EGFR is concomitantly expressed with its ligands, EGF or tumor growth

---

This work was presented at the 19th Bristol-Myers Squibb Nagoya International Cancer Treatment Symposium, "State of the Arts for Digestive Organs", 14–15 November 2003, Nagoya, Japan.

---

H. Q. Xiong  
Department of Gastrointestinal Medical Oncology, Unit 426,  
University of Texas MD Anderson Cancer Center,  
1515 Holcombe Boulevard, Houston, TX 77030, USA  
E-mail: qxiong@mdanderson.org  
Tel.: +1-713-7922828  
Fax: +1-713-7451163

**Table 1** Molecular targets and novel agents in pancreatic cancer

Targets	Frequency (%) <sup>a</sup>	Novel agents
Receptor tyrosine kinases		
EGFR	90	mAbs: cetuximab, ABX-EGF, EMD 72000 TKIs: gefitinib (ZD1839, Iressa), erlotinib (OSI-774, Tarceva), EKB-569 Herceptin, CI-1033
HER2/Neu	10	
Ras-Raf-MEK-ERK signaling pathways		
Ras	90	FTIs: R115777, SCH66336, BMS-214662
Raf		Bay 43-9006
MEK		CI-1040
PI3 K/Akt pathways		
Akt		17-AAG (nonspecific)
mTOR		CCI-779, RAD001
NF- $\kappa$ B	67	Curcumin (nonspecific), bortezomib (PS-341, VELCADE) (nonspecific)
Other molecular targets		
COX-2	75	Celecoxib, rofecoxib
LOX		LY293111
IL-8	70	ABX-IL8

<sup>a</sup>Mutation/expression rate in pancreatic cancer

factor  $\alpha$ , and the increased expression of ligand and receptor forms an autocrine loop that constantly stimulates cell proliferation [35, 43]. Finally, in pancreatic cancer, expression of EGFR and its ligands is associated with a poor prognosis [22, 72].

After being synthesized in the cytoplasm, the Ras proteins undergo a series of post-translational modifications culminating in membrane association. The first of these processing steps is farnesylation near the carboxy-terminal cysteine residue by farnesyl protein transferase. *Ras* carries a mutation at codon 12 (*K-ras*) in more than 90% of pancreatic cancer specimens [4]. The *K-ras* mutation results in constitutive activation of an intracellular signaling pathway, leading to cellular proliferation and thus conferring transforming properties onto cells containing point mutations in this gene. *Ras* mutation is considered an early genetic event in the development of pancreatic cancer, but *ras* mutation is not associated with tumor stage or prognosis, indicating that the *K-ras* oncogene may be related to the initiation of carcinogenesis, but is not linked to malignant potential or promotion of human pancreatic cancer [42].

One of the key downstream targets of the Ras family is phosphoinositol 3 kinase (PI3 K), a heterodimer consisting of a p85 regulatory subunit and a p110 catalytic subunit [14]. Activation of PI3 K can occur by binding of the p85 subunit to activated receptor tyrosine kinases or by binding of the p110 subunit to constitutively active Ras. Preclinical studies have shown that inhibitors of PI3 K, such as wortmannin and LY294002, induce dose-dependent apoptosis of pancreatic cancer cells that display constitutive Akt activity *in vitro*, and inhibit tumor growth of pancreatic cancer xenografts [10]. Activation of PI3 K is implicated in pancreatic cancer resistance to apoptosis induced by chemotherapeutic or molecular targeting agents, as several studies have demonstrated that treatment with the PI3 K inhibitors substantially enhances apoptosis induced by

gemcitabine in a concentration-dependent manner [11, 12, 45, 46]. Furthermore, Western blotting has shown that the reduction of phosphorylated Akt levels correlates with the enhancement of gemcitabine-induced apoptosis, suggesting that the PI3 K/Akt pathway plays a significant role in mediating drug resistance in human pancreatic cancer cells. Inhibition of the PI3 K/Akt signaling pathway also sensitizes pancreatic tumor cells to nonsteroidal antiinflammatory drugs (NSAIDs) such as sulindac [76]. Since the PI3 K/Akt signaling pathway plays an important role in tumor growth and resistance to apoptosis, it is a reasonable target for novel drug development.

The tumor suppressor gene PTEN is known to play a major role in embryonic development, cell migration and apoptosis [71]. PTEN acts as a lipid phosphatase that regulates major signal transduction pathways and effectively terminates PI3 K-mediated signaling [48]. PTEN mutation, which occurs frequently in many solid tumors, is associated with constitutive activation of the PI3 K/Akt pathway, resulting in tumors that are generally resistant to apoptosis. In pancreatic cancer, PTEN is not mutated but functionally abrogated through loss of expression. It was found that over 60% of pancreatic cancer cell lines and tumor tissues had decreased or loss of expression of PTEN (S. Reddy, personal communication). PTEN status in tumor cells has been implicated as an important predictor of sensitivity to sirolimus (formerly known as rapamycin) analogs [55] that inhibit the mammalian target of rapamycin (mTOR), a downstream effector of Akt.

Nuclear factor  $\kappa$ B (NF- $\kappa$ B) is a transcription factor that predominantly exists as p65 (RelA)/p50 heterodimer [27]. In most cells, NF- $\kappa$ B is sequestered in the cytoplasm in an inactive form through a noncovalent association with the inhibitor I $\kappa$ B $\alpha$ . This association masks the nuclear localization signal of NF- $\kappa$ B and thus prevents NF- $\kappa$ B nuclear translocation. Wang and

colleagues reported that RelA, the p65 subunit of NF- $\kappa$ B, was constitutively activated in approximately 67% of pancreatic adenocarcinomas, but not in healthy pancreatic tissues, and I $\kappa$ B $\alpha$  was overexpressed in human pancreatic tumor tissues and cell lines [67]. These data are consistent with the possibility that RelA is constitutively activated by the upstream signaling pathway, such as Ras, in pancreatic tumor cells. NF- $\kappa$ B may play an important role in tumor resistance to apoptosis induced by cytotoxic agents [6, 23]. Arlt and coworkers reported that pancreatic cancer cells resistant to gemcitabine exhibit a high basal NF- $\kappa$ B activity. Furthermore, gemcitabine showed a dose-dependent induction of NF- $\kappa$ B. Suppression of NF- $\kappa$ B by pharmacological or genetic approaches diminished the resistance against gemcitabine. NF- $\kappa$ B is, therefore, a potential target of novel drug development [6, 19, 25, 53].

Cyclooxygenases are enzymes that catalyze the conversion of arachidonic acid to various prostaglandins and thromboxanes and have a key role in inflammation and regulation of physiological functions. Two isoforms of the enzyme have been identified: cyclooxygenase-1 (COX-1) and cyclooxygenase-2 (COX-2). COX-1 supplies tissues with the prostaglandins required to maintain physiological organ function, such as cytoprotection of the gastric mucosa and regulation of renal blood flow. In contrast, COX-2 behaves as an immediate early gene and is subject to rapid regulation at the transcriptional level in response to inflammation, growth factors, cytokines and tumor promoters. Studies within the past decades have provided evidence that COX-2, but not COX-1, is induced in several types of human cancers, including pancreatic cancer. It has been reported that COX-2 expression can be detected in approximately 75% of pancreatic adenocarcinomas, with 50% of samples characterized as having high expression relative to adjacent normal tissue [26, 40, 64]. Numerous studies have shown that the NSAIDs and COX-2-specific inhibitors, celecoxib and rofecoxib, inhibit cell growth in both COX-2-positive and COX-2-negative pancreatic tumor cell lines in vitro and in vivo [19, 40, 63, 74, 75]. However, suppression of cell growth was significantly greater in the COX-2-expressing cell lines compared with COX-2-negative cell lines. The antiproliferative effect of COX-2 inhibitors is the consequence of their effect on cell-cycle arrest. Studies have indicated that treatment with COX-2 inhibitors results in the accumulation of proteins that are involved in arresting the cell cycle at G<sub>1</sub>, including p27, p21/WAF1 and others [63, 75]. The antiangiogenic property of COX-2 inhibitors may also contribute to their antitumor activity [34, 37]. Leahy and colleagues reported that celecoxib inhibited proliferation and promoted apoptosis of endothelial cells in celecoxib-treated xenograft tumors [34]. A recent study has demonstrated that celecoxib and NSAIDs directly induce apoptosis of endothelial cells in a pancreatic cancer xenograft model, but not through decreased production of angiogenic factors (Raut C. et al., submitted for publication). Thus, the antitumor activity of celecoxib

may be attributable, at least in part, to a direct effect on endothelial cells. These data suggest that COX-2 may play an important role in pancreatic tumorigenesis and therefore may be a promising chemotherapeutic target for the treatment of pancreatic cancer.

Lipoxygenase (LOX) pathways metabolize arachidonic acid to produce several potent biological mediators, including leukotriene B<sub>4</sub> (LTB<sub>4</sub>), the peptidoleukotrienes and hydroperoxyeicosatetraenoic acids (HETE) [21]. LOX, like COX, may play an important role in pancreatic carcinogenesis [20]. First, expression of LOX is upregulated in pancreatic cancer [18, 29]. Second, LOXs and their products enhance proliferation of pancreatic cancer cells [2, 19, 29, 62]. It was reported that expression of 5-LOX and the LTB<sub>4</sub> receptor in human pancreatic cancer tissues is markedly elevated and functions as an autocrine loop that stimulates proliferation of pancreatic cancer [29]. Third, inhibitors of LOX, such as LY293111, block growth of pancreatic cancer in vitro and in vivo [62].

Another important genetic event that is a potential therapeutic target is the overexpression of interleukin 8 (IL-8). IL-8 is a pleiotropic cytokine that plays an important role in inflammation, proliferation and angiogenesis, which are all significantly relevant to tumor growth and metastasis. IL-8 is overexpressed in about 70% of human pancreatic cancer cell lines [33]. Studies of pancreatic cancer cell lines that have different expression levels of IL-8 have indicated that increased IL-8 expression correlates with fast tumor growth, increased angiogenesis and more frequent metastasis [33, 57]. Moreover, abrogation of IL-8 expression using the antisense approach effectively suppresses tumor growth rate, angiogenesis and metastasis. Furthermore, preclinical studies have demonstrated that EGFR inhibitors suppress tumor growth in xenograft models in part through antiangiogenesis, including inhibition of IL-8 expression. A fully human anti-IL-8 antibody, ABX-IL8, is available and has shown antiangiogenic activity and activity against melanoma and bladder cancer, but there has been no further development for pancreatic cancer [31, 39].

Detailed understanding of these molecular events in pancreatic carcinogenesis has provided a foundation for the development of novel therapies targeting these molecular events. The rest of this review focuses on discussion of novel therapies that are in clinical development (Table 1). There are many other genetic changes, especially in tumor suppressor genes such as p53, p16, SMAD, etc. These genetic changes are important in pancreatic cancer development, but they are not easily "druggable" and are not discussed here.

### **EGFR as a therapeutic target**

Since EGFR plays a central role in controlling the activity of the Ras-Raf-MEK-ERK signaling pathway, great effort has been spent on developing strategies targeting EGFR. Monoclonal antibodies (mAbs) against the extracellular domain have proven a successful

approach to inhibit EGFR. The advent of hybridoma technology in the mid-1970s enabled fast mass production of mAbs. Using such a technique, Mendelsohn and his colleagues generated M225, a mAb that blocks activation of EGFR through specific binding to the extracellular domain of EGFR and competes with the natural ligands [51]. In vitro and in vivo studies have demonstrated that blockade of EGFR results in the arrest of cell-cycle progression and tumor growth inhibition [69]. Cetuximab (IMC-C225) is a chimeric mAb generated from fusion of the variable region of the murine M225 and the human IgG1 constant region. The resultant antibody retains high affinity for and specificity to EGFR and reduces immunogenicity. Preclinical studies have demonstrated that IMC-C225 alone effectively inhibits the proliferation of a variety of EGFR expressing cells in vitro and tumor growth in xenograft models [28].

Subsequent molecular and genetic engineering studies have further improved murine mAbs by introducing human immunoglobulin genes into mice by transgenic technology (XenoMouse) [17, 38, 49]. ABX-EGF is a human antibody against EGFR that is generated using the XenoMouse technique and is in the early phase of clinical development [73]. Another humanized anti-EGFR antibody, EMD 72000, entered clinical development recently [32]. The advantage of using an antibody as a drug is its high affinity and predictable specificity. EGFR undergoes internalization and subsequent degradation upon ligand binding. MAb binds to EGFR in a way that mimics ligands, causing EGFR degradation, and therefore further enhances their inhibitory effects. Although several mAbs against EGFR are available, only cetuximab has been studied extensively in pancreatic cancer models. Cetuximab, as expected, can effectively block EGFR autophosphorylation in vitro and in vivo. An additive inhibitory effect was observed when cetuximab was combined with either gemcitabine or fluorouracil [13, 47].

Histological study of tumor specimens obtained from mice that received treatment with either cetuximab or cetuximab in combination with gemcitabine revealed that cetuximab induces apoptosis and suppresses proliferation of tumor cells [12]. More interesting was the finding that EGFR-targeted therapies induce apoptosis of endothelial cells, which are not thought to be direct targets of EGFR inhibition. In addition, decreased microvascular densities, as well as production of VEGF and IL-8, were observed in the tumor in response to cetuximab. These data suggested that, besides antiproliferative activity, antiangiogenic activity contributes significantly to the antitumor effects of EGFR inhibitors. A phase II trial of cetuximab in combination with gemcitabine for advanced pancreatic cancer was reported recently [1]. In that trial, 41 patients whose tumors expressed EGFR were treated with cetuximab at 400 mg/m<sup>2</sup> for the first dose and then 250 mg/m<sup>2</sup> weekly plus gemcitabine at standard dose and schedule. A partial response rate of 12.2% and disease control (sum of partial response and stable disease) of 75.6% were

observed. The median survival duration was 7.1 months and the 1-year survival rate was 31.7%. This encouraging activity prompted the proposal of a phase III trial comparing gemcitabine to gemcitabine plus cetuximab by the US Southwest Oncology Group.

Despite the fact that protein tyrosine domains share significant amino acid sequence homology and a highly conserved core structure, the ATP-binding site has been proven to be an exciting target for drug design. Many small molecules of tyrosine kinase inhibitors (TKIs) have been synthesized and are in different phases of clinical development, including gefitinib (ZD1839, Iressa), erlotinib (OSI-774, Tarceva) and EKB-569. The potential advantages of TKIs include the potentially easy production of large quantities and the fact that one molecule can potentially inhibit a family of tyrosine kinases that share a similar structure. At present, TKIs have been studied in combination with gemcitabine for advanced pancreatic cancer. The results of a phase I trial of EKB-569 in combination with gemcitabine for advanced pancreatic cancer were presented at the 2003 American Society of Clinical Oncology (ASCO) annual meeting [41]. The dose-limiting toxicities were grade 3 diarrhea and elevation of transaminases. The maximum tolerated dose (MTD) for the combination was EKB-569 25 mg plus gemcitabine 750 mg/m<sup>2</sup>. Twenty patients were treated at the MTD, allowing adequate assessment of the antitumor activity of this combination. The National Cancer Institute of Canada is conducting a randomized placebo-controlled phase III study of erlotinib plus gemcitabine versus gemcitabine in patients with advanced pancreatic cancer. The accrual has been completed and the results are eagerly awaited.

---

### **Ras-Raf-MEK signaling pathways as therapeutic targets**

Ras proteins undergo serial steps of modification. The key step is farnesylation by the enzyme farnesyl-protein transferase (FPTase), which adds the 15-carbon farnesyl isoprenoid to a cysteine residue four amino acids from the COOH-terminus of Ras. Oncogenic forms of Ras, as well as wildtype Ras, require this COOH-terminal prenylation for their biological and/or transforming functions. These findings provided the impetus to develop farnesyl transferase inhibitors (FTIs) as a means of targeting Ras for the treatment of cancer [54]. FTIs inhibit the growth of both normal cells and cancer cells containing either wildtype or mutant forms of *ras*, although cells transformed with oncogenic *H-ras* tend to be much more responsive than cells harboring wildtype *ras* or *K-ras*, or *N-ras* [24]. Several FTIs are in clinical development, including R115777 (Zarnestra) [16, 36, 66], SCH66336 (Lonafarnib, Sarasar) [61] and BMS-214662.

R115777, an oral agent, is a selective nonpeptidomimetic inhibitor of FPTase. Preclinical studies demonstrated that R115777 competitively inhibits farnesylation of lamin B and K-RasB peptides at nanomolar concentrations while inhibiting proliferation

of pancreatic cancer cell lines and xenografts. However, the results of phase II and III clinical studies have been disappointing. A phase II trial of R115777 as initial therapy for patients with advanced pancreatic cancer in which 20 patients were enrolled yielded no objective responses. Median survival time was 19.7 weeks and the estimated 6-month survival rate was 25%. In this study, inhibition of FPTase activity in peripheral blood mononuclear cells was measured; a partial inhibition of FPTase activity was observed despite lack of clinical activity [16]. In a double-blind phase III trial, Van Cutsem and colleagues [66] tested R115777 in combination with gemcitabine against gemcitabine plus placebo. The median overall survival times were 193 and 182 days for patients in the R115777 and placebo arms, respectively. There were no differences in the rate of progression-free survival, 6-month survival, or 1-year survival.

The disappointing clinical results raise the question of whether Ras is a valid target for pancreatic cancer. Ras is a key member of the signaling pathways that regulate critical cellular functions, and *ras* mutation has the ability to transform normal cells into neoplastic phenotype and occurs frequently in human cancers. Therefore, Ras appears to be an important target for drug development. However, in pancreatic cancer, *ras* mutation occurs early, indicating its role in the early development of pancreatic cancer. In contrast, its role in established pancreatic cancer is not clear. Moreover, more than 90% of pancreatic cancers bearing *K-ras* have been demonstrated to be less sensitive to FTIs. In addition, Ras isoforms and oncogenic Ras can be geranylgeranylated (by geranylgeranyl transferase I), an alternative lipidation that can substitute for farnesylation. Finally, advanced pancreatic cancer, like many solid tumors, gains multiple survival advantages that may compensate for ras inhibition.

Raf is a signaling protein downstream of Ras. Ras activates the Raf-MEK-ERK pathway by first localizing Raf to the plasma membrane, where Raf initiates a mitogenic kinase cascade that leads to cell proliferation. Studies with dominant-negative mutants and antisense molecules suggest that inhibition of Raf kinase is an important target for cancer therapy. Bay 43-9006 is a small-molecule inhibitor that was designed specifically to target Raf kinase. Preclinical studies indicate that Bay 43-9006 is a potent inhibitor of Raf kinase in vitro and in vivo, with significant dose-dependent antitumor activity. Pancreatic cancer should be a good tumor type for the clinical investigation of Bay 43-9006, since the Ras-Raf-MEK-ERK pathway is constitutively activated. A phase I study of Bay 43-9006 in combination with gemcitabine has determined that the recommended phase II dose is BAY 43-9006 400 mg twice daily and gemcitabine 1000 mg/m<sup>2</sup> weekly  $\times 7$ , followed by 1 week rest, then weekly  $\times 3$  every 4 weeks [58]. The efficacy of this combination in pancreatic cancer is being evaluated at the recommended phase II dose. An ongoing phase II study of Bay 43-9006 for patients with solid tumors,

including colorectal cancer, renal cell carcinoma, malignant melanoma, pancreatic cancer and other tumor types, will also provide an initial assessment of the antitumor efficacy of Bay 43-9006 as a single agent.

CI-1040 is an oral, highly selective, small-molecule inhibitor of the dual specificity kinase MEK1/2. Preclinical antitumor activity has been demonstrated in a pancreatic cancer xenograft. A phase II study of CI-1040 was conducted for patients with advanced small-cell lung cancer, breast cancer, colon cancer, and pancreatic cancer [68]. The study employed a two-stage design: in each tumor type, either one objective response (CR/PR) or four clinical benefit responses (CBR = CR/PR/SD) in stage 1 ( $n = 13$ ) would trigger stage 2 ( $n = 30$ ) enrollment, resulting in a total of 43 patients for each of the four tumor types. Unfortunately, the study did not advance to stage 2 due to limited antitumor activity.

---

### Antiapoptotic signaling pathways as targets

Two potential antiapoptotic signal transduction pathways have been linked to chemoresistance of pancreatic carcinoma cell lines: the PI3 K/Akt pathway and the NF- $\kappa$ B pathway. Besides the growth-promoting potential of the PI3 K/Akt pathway, its antiapoptotic properties are closely linked to the resistance of cancer cells to a broad spectrum of apoptotic stimuli. Therefore, the PI3 K/Akt pathway is an important target for drug development, but no specific agents have entered clinical development yet. Recent studies have indicated that Akt is a client protein of heat shock protein 90 (Hsp90), which can be inhibited by the benzoquinone ansamycin antibiotics (BA), herbimycin, geldanamycin and 17-allylamino-17-demethoxygeldanamycin (17-AAG). It was reported that occupancy of the Hsp90 pocket by ansamycins results in a reduction in Akt half-life and protein expression secondary to Akt ubiquitination and proteasomal degradation [9]. 17-AAG has undergone phase I clinical investigations [8]. It would be interesting to test 17-AAG as monotherapy or in combination for pancreatic cancer.

mTOR, a downstream molecule of Akt, is located predominantly in the nuclear fraction of both neoplastic and normal cells [77]. mTOR activation triggers resting cells to increase the translation of a subset of mRNAs whose proteins are required for cell-cycle progression from G<sub>1</sub> to S phase. CCI-779, an ester of the macrocyclic immunosuppressive agent sirolimus, reacts with the ubiquitous intracellular FK506-binding protein 12 (FKBP12) [7, 26]. The CCI-779/FKBP12 complex is a potent inhibitor of the highly conserved kinase mTOR. Inhibition of mTOR leads to suppression of several downstream signaling effectors, including the ribosomal subunit p70<sup>S6</sup> kinase and the eukaryotic initiation factor 4 binding protein 1 (4E-BP1) [11]. The extent of phosphorylation of these two downstream proteins (p70<sup>S6</sup> kinase and 4E-BP1) may therefore serve as indicators of CCI-779 biologic activity in vivo. Clinical trials have

shown that CCI-779 is well tolerated at a variety of doses and schedules. Furthermore, antitumor activity has been observed in breast cancer and renal cell carcinoma [15, 30]. Pancreatic cancer is hypothesized to be sensitive to CCI-779 since PTEN expression is either undetectable or significantly decreased. A phase II trial of CCI-779 for advanced pancreatic cancer is being planned.

NF- $\kappa$ B is an important target for therapeutic intervention because of its role in promoting cancer cell growth and developing resistance to apoptosis. However, no NF- $\kappa$ B-specific agents are available for clinical development. The activity of NF- $\kappa$ B is regulated by I $\kappa$ B $\alpha$ , which is degraded by the ubiquitin-proteasome pathway. Bortezomib (PS-341, VELCADE) is a dipeptide boronate antagonist of the proteasome and blocks I $\kappa$ B $\alpha$  degradation and subsequent NF- $\kappa$ B activation. In vitro and in vivo studies have demonstrated that bortezomib exhibits a wide range of activity, from extremely sensitive to resistant, against pancreatic cancers. However, these effects do not correlate with differential inhibition of NF- $\kappa$ B activation, indicating that inhibition of NF- $\kappa$ B per se is not always sufficient to induce apoptosis [44]. Ryan and coworkers [50] combined bortezomib (administered twice weekly for 2 weeks) with gemcitabine (given on days 1 and 8) in a phase I clinical study of advanced solid tumors. The combination of gemcitabine at a dose of 1000 mg/m<sup>2</sup> and bortezomib at a dose of 1.0 mg/m<sup>2</sup> showed good tolerability. However, the efficacy of bortezomib for the treatment of pancreatic cancer remains to be elucidated.

Curcumin (diferuloylmethane), a polyphenol derived from the plant *Curcuma longa*, has demonstrated activity against a wide range of tumor cells in vitro and in vivo through inhibition of NF- $\kappa$ B and a variety of other molecules involved in cell growth [3]. Despite its diverse effect against many molecules and promising antitumor activity in animal studies, the clinical development of curcumin has lagged behind. A phase I study of curcumin revealed that curcumin extract was well tolerated and dose-limiting toxicity was not reached, but bioavailability was low [56].

### COX and LOX as targets

Both COX and LOX are implicated in pancreatic carcinogenesis, although the role of LOX has been less defined than the role of COX. The NSAIDs have long demonstrated antitumor activity. The development of COX-2-specific inhibitors (for example, celecoxib and rofecoxib), drugs that maintain their antiinflammatory properties while preserving the biosynthesis of protective prostaglandins, further raised interest in targeting COX-2 in cancer therapy. A phase I trial of gemcitabine in combination with celecoxib was conducted to study potential drug interactions [70]. The results showed that there was no alteration of gemcitabine conversion to its active metabolite with the addition of celecoxib. The

study determined that the doses recommended for further study were gemcitabine 650 mg/m<sup>2</sup> over a 65-min infusion for three consecutive weeks with 1 week rest and celecoxib 400 mg twice daily. The preliminary results of a phase II trial of gemcitabine plus celecoxib were presented at the ASCO 2003 meeting [59]. The study enrolled 20 patients with advanced pancreatic cancer; three patients had partial responses and four had stable disease.

LY293111 is a biphenyl substituted diaryl ether carboxylic acid originally discovered as an LTB<sub>4</sub> receptor antagonist. Phase I studies of LY293111 as a single agent or in combination with gemcitabine showed that LY293111 was well tolerated, with diarrhea as the most frequently reported adverse event [52, 60]. There was no interaction between LY293111 and gemcitabine, and gemcitabine could be administered at full dose when combined with LY293111. A randomized, double-blind phase II trial of gemcitabine plus LY293111 versus placebo for patients with advanced pancreatic cancer has completed accrual and the results are eagerly awaited.

### Conclusion

The advance of molecular biology has led to the elucidation of molecular events important for pancreatic carcinogenesis and provided a foundation for the development of novel therapies. Several classes of agents, such as, TKIs, FTIs, and COX-2 and LOX inhibitors, have shown promising activity in the preclinical setting, but the clinical data so far are less impressive. Cetuximab, a monoclonal antibody against EGFR, in combination with gemcitabine has shown promising antitumor activity for advanced pancreatic cancer, and a further phase III randomized trial of this combination versus gemcitabine is being planned. Meanwhile a phase III trial of gemcitabine plus erlotinib versus placebo has completed accrual. At the present time, gemcitabine remains the standard chemotherapy for advanced pancreatic cancer.

### References

1. Abbruzzese J, Rosenberg A, Xiong Q, LoBuglio A, Schmidt W, Wolff R, Needle M, Waksal H (2001) A phase II study of anti-epidermal growth factor receptor (EGFR) antibody cetuximab (IMC-C225) in combination with gemcitabine in patients with advanced pancreatic cancer (abstract 518). Proc Am Soc Clin Oncol 20
2. Adrian TE, Ding XZ, Tong WG (2001) The mitogenic effect of 5-lipoxygenase metabolite, 5-HETE on pancreatic cancer cell proliferation through activation of multiple signaling pathways. FASEB J 15:A1163
3. Aggarwal BB, Kumar A, Bharti AC (2003) Anticancer potential of curcumin: preclinical and clinical studies. Anticancer Res 23(1A):363
4. Almoguera C, Shibata D, Forrester K, Martin J, Arnheim N, Perucho M (1988) Most human carcinomas of the exocrine pancreas contain mutant c-K-ras genes. Cell 53:549

5. Alroy I, Yarden Y (1988) The ErbB signaling network in embryogenesis and oncogenesis: signal diversification through combinatorial ligand-receptor interactions. *FEBS Lett* 410:83
6. Arlt A, Gehrz A, Muerkoster S, Vorndamm J, Kruse ML, Folsch UR, Schafer H (2003) Role of NF-kappaB and Akt/PI3 K in the resistance of pancreatic carcinoma cell lines against gemcitabine-induced cell death. *Oncogene* 22:3243
7. Armistead D, Harding MW (1993) Immunophilins and immunosuppressive drug action. *Ann Rev Med Chem* 28:207
8. Banerji U, O'Donnell A, Scurr M, Benson C, Stapleton S, Raynaud F, Clarke F, Turner A, Workman P, Judson I, Block E (2003) A pharmacokinetically (PK)-pharmacodynamically (PD) guided phase I trial of the heat shock protein 90 (HSP90) inhibitor 17-allylamino,17-demethoxygeldanamycin (17AAG) (abstract 797). *Proc Am Soc Clin Oncol* 22:199
9. Basso AD, Solit DB, Chiosis G, Giri B, Tschlis P, Rosen N (2002) Akt forms an intracellular complex with heat shock protein 90 (Hsp90) and Cdc37 and is destabilized by inhibitors of Hsp90 function. *J Biol Chem* 277:39858
10. Bondar VM, Sweeney-Gotsch B, Andreeff M, Mills GB, McConkey DJ (2002) Inhibition of the phosphatidylinositol 3'-kinase-AKT pathway induces apoptosis in pancreatic carcinoma cells in vitro and in vivo. *Mol Cancer Ther* 1:989
11. Brown EJ, Albers MW, Shin TB, Khikawa K, Keith CT, Lane WS, Shreiber SL (1994) A mammalian protein targeted by G<sub>i</sub>-arresting rapamycin-receptor complex. *Nature* 369:756
12. Bruns CJ, Harbison MT, Davis DW, Portera CA, Tsan R, McConkey DJ, Evans DB, Abbruzzese JL, Hicklin DJ, Radinsky R (2000) Epidermal growth factor receptor blockade with C225 plus gemcitabine results in regression of human pancreatic carcinoma growing orthotopically in nude mice by antiangiogenic mechanisms. *Clin Cancer Res* 5:1936
13. Buchsbaum DJ, Bonner JA, Grizzle WE, Stackhouse MA, Carpenter M, Hicklin DJ, Bohlen P, Raisch KP (2002) Treatment of pancreatic cancer xenografts with Erbitux (IMC-C225) anti-EGFR antibody, gemcitabine, and radiation. *Int J Radiat Oncol Biol Phys* 4:1180
14. Cantley LC (2003) The phosphoinositide 3-kinase pathway. *Science* 5573:1655
15. Chan S, Scheulen ME, Johnston S, Mross K, Piccart M, Hess D, Bouxin N, Azli N (2003) Phase 2 study of two dose levels of CCI-779 in locally advanced or metastatic breast cancer (MBC) failing prior anthracycline and/or taxane regimens (abstract 774). *Proc Am Soc Clin Oncol* 22:193
16. Cohen SJ, Ho L, Ranganathan S, Abbruzzese JL, Alpaugh RK, Beard M, Lewis NL, McLaughlin S, Rogatko A, Perez-Ruix JJ, Thistle AM, Verhaeghe T, Wang H, Weiner LM, Wright JJ, Hudes GR, Meropol NJ (2003) Phase II and pharmacodynamic study of the farnesyltransferase inhibitor R115777 as initial therapy in patients with metastatic pancreatic adenocarcinoma. *J Clin Oncol* 21:1301
17. Davis CG, Gallo ML, Corvalan JRF (2003) Transgenic mice as a source of fully human antibodies for the treatment of cancer. *Cancer Metastasis Rev* 18:421
18. Ding XZ, Iversen P, Cluck MW, Knezetic JA, Adrian TE (1999) Lipoxigenase inhibitors abolish proliferation of human pancreatic cancer cells. *Biochem Biophys Res Commun* 261:218
19. Ding XZ, Tong WG, Adrian TE (2000) Blockade of cyclooxygenase-2 inhibits proliferation and induces apoptosis in human pancreatic cancer cells. *Anticancer Res* 4:2625
20. Ding XZ, Hennig R, Adrian TE (2003) Lipoxigenase and cyclooxygenase metabolism: new insights in treatment and chemoprevention of pancreatic cancer. *Mol Cancer* 1:10
21. Ding XZ, Tong WG, Adrian TE (2003) 12-Lipoxigenase metabolite, 12(S)-HETE stimulates pancreatic cancer cell proliferation. Involvement of tyrosine phosphorylation and MEK/ERK activation. *Int J Cancer* 94:630
22. Dong M, Nio Y, Guo KJ, Tamura K, Tian YL, Dong YT (1998) Epidermal growth factor and its receptor as prognostic indicators in Chinese patients with pancreatic cancer. *Anticancer Res* 18:4613
23. Dong QG, Scwabas GM, Fujioka S, Schmidt C, Peng B, Wu T, Tsao MS, Evans DB, Abbruzzese JL, McDonnell TJ, Chiao PJ (2003) The function of multiple IkappaB:NF-kappaB complexes in the resistance of cancer cells to Taxol-induced apoptosis. *Oncogene* 21:6510
24. End DW, Smets G, Todd AV, Applegate TL, Fuery CJ, Angibaud P, Venet M, Sanz G, Poignet H, Skrzat S, Devine A, Wouters W, Bowden C (2001) Characterization of the antitumor effects of the selective farnesyl protein transferase inhibitor R115777 in vivo and in vitro. *Cancer Res* 61:131
25. Fujioka S, Scwabas GM, Schmidt C, Frederick WA, Dong QG, Abbruzzese JL, Evans DB, Baker C, Chiao PJ (2003) Function of nuclear factor kappaB in pancreatic cancer metastasis. *Clin Cancer Res* 9:346
26. Geoerger B, Kerr K, Tang CB, Fung KM, Powell B, Sutton LN, Phillips PC, Janss AJ (2001) Antitumor activity of the rapamycin analogue CCI-779 in human primitive neuroectodermal tumor/medulloblastoma models as single agent and in combination chemotherapy. *Cancer Res* 61:1527
27. Gilmore TD, Koedood M, Piffat KA, White D (1997) Rel/NF-kappaB/IkappaB proteins and cancer. *Oncogene* 14:1367
28. Goldstein NI, Prewett M, Zuklys K, Rockwell P, Mendelsohn J (1995) Biological efficacy of a chimeric antibody to the epidermal growth factor receptor in a human tumor xenograft model. *Clin Cancer Res* 1:1311
29. Hennig R, Ding XZ, Tong WG, Schneider MB, Standop J, Friess H, Buchler MW, Pour PM, Adrian TE (2002) 5-Lipoxygenase and leukotriene B<sub>4</sub> receptor are expressed in human pancreatic cancers but not in pancreatic ducts in normal tissue. *Am J Pathol* 161:421
30. Hidalgo M, Atkins MB, Stadler WM, Logan T, Dutcher JP, Hudes G, Marshall B, Liou SH, Dukart G (2003) A randomized double-blind phase 2 study of intravenous (IV) CCI-779 administered weekly to patients with advanced renal cell carcinoma (RCC): prognostic factor analysis (abstract 804). *Proc Am Soc Clin Oncol* 22:201
31. Huang S, Mills L, Mian B, Tellez C, McCarty M, Yang XD, Gudas JM, Bar-Eli M (2002) Fully humanized neutralizing antibodies to interleukin-8 (ABX-IL8) inhibit angiogenesis, tumor growth, and metastasis of human melanoma. *Am J Pathol* 161:125
32. Kollmannsberger C, Schittenhelm M, Honecker F, Rosen O, Tillner J, Kanz L, Bokemeyer C (2003) Epidermal growth factor receptor (EGFR) antibody EMD 72000 in combination with paclitaxel (P) in patients (pts) with EGFR-positive advanced non-small cell lung cancer (NSCLC): a phase-I study (abstract 2520). *Proc Am Soc Clin Oncol* 22:627
33. Le X, Shi Q, Wang B, Xiong Q, Qian C, Peng Z, Li XC, Tang H, Abbruzzese JL, Xie K (2000) Molecular regulation of constitutive expression of interleukin-8 in human pancreatic adenocarcinoma. *J Interferon Cytokine Res* 20:935
34. Leahy KM, Ornberg RL, Wang Y, Zweifel BS, Koki AT, Masferrer JL (2002) Cyclooxygenase-2 inhibition by celecoxib reduces proliferation and induces apoptosis in angiogenic endothelial cells in vivo. *Cancer Res* 62:625
35. Lemoine NR, Hughes CM, Barton CM, Poulsom R, Jeffery RE, Kloppel G, Hall PA, Gullick WJ (1992) The epidermal growth factor receptor in human pancreatic cancer. *J Pathol* 166:7
36. Macdonald JS, Chansky K, Whitehead R, Wade J, Giguere J, Abbruzzese JL (2002) A phase II study of farnesyl transferase inhibitor R115777 in pancreatic cancer: a Southwest Oncology Group (SWOG) (abstract 548). *Proc Am Soc Clin Oncol* 21
37. Masferrer JL, Leahy KM, Koki AT, Zweifel BS, Settle SL, Woerner BM, Edwards DA, Flickinger AG, Moore RJ, Seibert K (2000) Antiangiogenic and antitumor activities of cyclooxygenase-2 inhibitors. *Cancer Res* 60:1306
38. Mendez MJ, Green LL, Corvalan JRF, Jia XC, Maynard-Curie CE, Yang XD, Gallo ML, Louie DM, Lee DV, Erickson KL, Luna J, Roy CM, Abderrahim H, Kirschenbaum F, Noguchi M, Smith DH, Fukushima A, Hales JF, Klapholz S,

- Finer MH, Davis CG, Zsebo KM, Jakobovits A (1997) Functional transplant of megabase human immunoglobulin loci recapitulates human antibody response in mice. *Nat Genet* 15:146
39. Mian BM, Dinney CP, Bermejo CE, Sweeney P, Tellez C, Yang XD, Gudas JM, McConkey DJ, Bar-Eli M (2002) Fully human anti-interleukin 8 antibody inhibits tumor growth in orthotopic bladder cancer xenografts via down-regulation of matrix metalloproteases and nuclear factor-kappaB. *Clin Cancer Res* 9:3167
40. Molina MA, Sitja-Arnau M, Lemoine MG, Frazier ML, Sinicrope FA (1999) Increased cyclooxygenase-2 expression in human pancreatic carcinomas and cell lines: growth inhibition by nonsteroidal anti-inflammatory drugs. *Cancer Res* 59:4356
41. Morgan JA, Bukowski RM, Xiong H, Clark J, Zacharchuk C, Plazney D, Pelley R, Fuchs C (2003) Preliminary report of a phase I study of EKB-569, an irreversible inhibitor of the epidermal growth factor receptor (EGFR), given in combination with gemcitabine to patients with advanced pancreatic cancer (abstract 788). *Proc Am Soc Clin Oncol* 22:197
42. Motojima K, Urano T, Nagata Y, Shiku H, Tsunoda T, Kanematsu T (1991) Mutations in the Kirsten-ras oncogene are common but lack correlation with prognosis and tumor stage in human pancreatic carcinoma. *Am J Gastroenterol* 86:1784
43. Murphy LO, Cluck MW, Lovas S, Otvos F, Murphy RF, Schally AV, Permert J, Larsson J, Knezetic JA, Adrian TE (2001) Pancreatic cancer cells require an EGF receptor-mediated autocrine pathway for proliferation in serum-free conditions. *Br J Cancer* 84:926
44. Nawrocki ST, Bruns CJ, Harbison MT, Bold RJ, Gotsch BS, Abbruzzese JL, Elliott P, Adams J, McConkey DJ (2002) Effects of the proteasome inhibitor PS-341 on apoptosis and angiogenesis in orthotopic human pancreatic tumor xenografts. *Mol Cancer Ther* 1:243
45. Ng SS, Tsao MS, Chow S, Hedley DW (2000) Inhibition of phosphatidylinositol 3-kinase enhances gemcitabine-induced apoptosis in human pancreatic cancer cells. *Cancer Res* 60:5451
46. Ng SS, Tsao MS, Nicklee T, Hedley DW (2001) Wortmannin inhibits pkb/akt phosphorylation and promotes gemcitabine antitumor activity in orthotopic human pancreatic cancer xenografts in immunodeficient mice. *Clin Cancer Res* 7:3269
47. Overholser JP, Prewett MC, Hooper AT, Waksal HW, Hicklin DJ (2000) Epidermal growth factor receptor blockade by antibody IMC-C225 inhibits growth of a human pancreatic carcinoma xenograft in nude mice. *Cancer* 89:74
48. Podsypanina K, Lee RT, Politis S, Hennessy I, Crane A, Puc J, Neshat M, Wang H, Yang L, Gibbons J, Frost P, Dreisbach V, Blenis J, Gaciong Z, Fisher P, Sawyers C, Hedrick-Ellenson L, Passons R (2001) An inhibitor of mTOR reduces neoplasia and normalizes p70/S6 kinase activity in *Pten*<sup>+/-</sup> mice. *Proc Natl Acad Sci U S A* 98:10320
49. Presta LG (1992) Antibody engineering. *Curr Opin Biotechnol* 3:394
50. Ryan DP, Elder JP, Winkelmann J, Lynch T, Supko J, Appleman LJ, Fidias P, Enzinger P, Zhu A, Kinchla N, Esseltine D, Baldwin A, Elliott P, Adams J, Kauffman M, Schenkein D, Cusack J (2002) Pharmacokinetic and pharmacodynamic phase I study of PS-341 and gemcitabine in patients with advanced solid tumors (abstract 379). *Proc Am Soc Clin Oncol* 21
51. Sato JD, Kawamoto T, Le AD, Mendelsohn J, Polikoff J, Sato GH (1983) Biological effects in vitro of monoclonal antibodies to human epidermal growth factor receptors. *Mol Biol Med* 1:511
52. Schwartz GK, Budman DR, Endres S, Welch M, O'Reilly E, Barile-Thiem B, Brail LH, De Alwis DP, Cleverly A, Weitzman A (2002) Phase I and pharmacokinetic study of LY293111, an orally available small molecule known to be an LTB4 receptor antagonist, 5-lipoxygenase inhibitor and peroxisome proliferator activated receptor-gamma agonist (PPAR $\gamma$ ) (abstract 343). *Proc Am Soc Clin Oncol* 21
53. Scialbas GM, Fujioka S, Schmidt C, Fan Z, Evans DB, Chiao PJ (2003) Restoring apoptosis in pancreatic cancer cells by targeting the nuclear factor-kappaB signaling pathway with the anti-epidermal growth factor antibody IMC-C225. *J Gastrointest Surg* 7:37
54. Sebt SM, Hamilton AD (2000) Farnesyltransferase and geranylgeranyltransferase I inhibitors in cancer therapy: important mechanistic and bench to bedside issues. *Expert Opin Investig Drugs* 9:2767
55. Sekulic A, Hudson CC, Homme JL, Yin P, Offerness DM, Karnitz LM, Abraham RT (2000) A direct linkage between the phosphoinositide 3-kinase-AKT signaling pathway and the mammalian target of rapamycin in mitogen-stimulated and transformed cells. *Cancer Res* 60:3504
56. Sharma RA, McLelland HR, Hill KA, Ireson CR, Euden SA, Manson MM, Pirmohamed M, Marnett LJ, Gescher AJ, Steward WP (2001) Pharmacodynamic and pharmacokinetic study of oral *Curcuma* extract in patients with colorectal cancer. *Clin Cancer Res* 7:1894
57. Shi Q, Abbruzzese JL, Huang S, Fidler IJ, Xiong Q, Xie K (1999) Constitutive and inducible interleukin 8 expression by hypoxia and acidosis renders human pancreatic cancer cells more tumorigenic and metastatic. *Clin Cancer Res* 5:3711
58. Siu L, Awada A, Takimoto CH, Moore MJ, Piccart M, Fiannder W, Lathia C, Petrenciu O (2003) Phase I study of oral raf-1 kinase inhibitor BAY 43-9006 with gemcitabine in patients with advanced solid tumors (abstract 828). *Proc Am Soc Clin Oncol* 22:207
59. Smith SE, Burris HA, Loehrer PJ, Sweeny C, Gordon M, O'Brien Y, Dragovich T (2003) Preliminary report of a phase II trial of gemcitabine combined with celecoxib for advanced pancreatic cancer (abstract 1502). *Proc Am Soc Clin Oncol* 22:374
60. Stevenson JP, Petrylak DP, K Feit K, Shelton G, Gallagher M, Haller DH, Sun WJ, Kindsfater S, Brail LH, de Alwis DP, Weitzman A Jr, O'Dwyer PJ (2002) Phase I and pharmacokinetic trial of LY293111 in combination with gemcitabine (abstract 2140). *Proc Am Soc Clin Oncol* 21
61. Theodore C, Geoffrois L, Vermorken JB, Caponigro F, Fiedler W, Chollet P, Ravaut A, Baron B, Lacombe D, Fumoleau P (2003) A phase II multicentre study of SCH66336 in combination with gemcitabine as second line treatment in patients with advanced/metastatic urothelial tract tumor (abstract 1667). *Proc Am Soc Clin Oncol* 22:415
62. Tong WG, Ding XZ, Hennig R, Witt RC, Standop J, Pour PM, Adrian TE (2002) Leukotriene B4 receptor antagonist LY293111 inhibits proliferation and induces apoptosis in human pancreatic cancer cells. *Clin Cancer Res* 8:3232
63. Tseng WW, Deganutti A, Chen MN, Saxton RE, Liu CD (2002) Selective cyclooxygenase-2 inhibitor rofecoxib (Vioxx) induces expression of cell cycle arrest genes and slows tumor growth in human pancreatic cancer. *J Gastrointest Surg* 6:838
64. Tucker ON, Dannenberg AJ, Yang EK, Zhang F, Teng L, Daly JM, Soslow RA, Masferrer JL, Woerner BM, Koki AT, Fahey TJ 3rd (1999) Cyclooxygenase-2 expression is up-regulated in human pancreatic cancer. *Cancer Res* 59:987
65. Ullrich A, Schlessinger J (1990) Signal transduction by receptors with tyrosine kinase activity. *Cell* 61:203
66. Van Cutsem E, Karasek P, Oettle H, Vervenne WL, Szawlowski A, Schoffski P, Post S, Neumann H, Safran H, Humblet Y, van de Velde H, Ma Y, Von Hoff D (2002) Phase III trial comparing gemcitabine + R115777 (Zarnestra) versus gemcitabine + placebo in advanced pancreatic cancer (PC) (abstract 517). *Proc Am Soc Clin Oncol* 21
67. Wang W, Abbruzzese JL, Evans DB, Larry L, Cleary KR, Chiao PJ (1999) The nuclear factor-kappa B RelA transcription factor is constitutively activated in human pancreatic adenocarcinoma cells. *Clin Cancer Res* 5:119
68. Waterhouse DM, Rinehart J, Adjei AA, Hecht JR, Natale RB, Lorusso PM, Asbury PC, Hamid O, Gulyas S, Meyer MB (2003) A phase 2 study of an oral MEK inhibitor, CI-1040, in patients with advanced nonsmall-cell lung, breast, colon, or pancreatic cancer (abstract 816). *Proc Am Soc Clin Oncol* 22:204



69. Wu X, Rubin M, Fan Z, De Blasio T, Soos T, Koff A, Mendelsohn J (1996) Involvement of p27<sup>kip1</sup> in G<sub>1</sub> arrest mediated by an anti-epidermal growth factor receptor monoclonal antibody. *Oncogene* 12:1397
70. Xiong HQ, Du M, Wolff RA, Lenzi R, Dumas P, Lassere Y, Plunkett W, Abbruzzese JL (2002) Pharmacology study of celecoxib in combination with gemcitabine for advanced pancreatic cancer (abstract 448). *Proc Am Soc Clin Oncol* 21
71. Yamada KM, Araki M (2001) Tumor suppressor PTEN: modulator of cell signaling, growth, migration and apoptosis. *J Cell Sci* 114:2375
72. Yamanaka Y, Friess H, Kobrin MS, Buchler M, Beger HG, Korc M (1993) Coexpression of epidermal growth factor receptor and ligands in human pancreatic cancer is associated with enhanced tumor aggressiveness. *Anticancer Res* 13:565
73. Yang XD, Jia XC, Corvalan JR, Wang P, Davis CG (2001) Development of ABX-EGF, a fully human anti-EGF receptor monoclonal antibody, for cancer therapy. *Crit Rev Oncol Hematol* 38:17
74. Yip-Schneider MT, Barnard DS, Billings SD, Cheng L, Heilman DK, Lin A, Marshall SJ, Crowell PL, Marshall MS, Sweeney CJ (2000) Cyclooxygenase-2 expression in human pancreatic adenocarcinomas. *Carcinogenesis* 21:139
75. Yip-Schneider MT, Sweeney CJ, Jung SH, Crowell PL, Marshall MS (2001) Cell cycle effects of nonsteroidal anti-inflammatory drugs and enhanced growth inhibition in combination with gemcitabine in pancreatic carcinoma cells. *J Pharmacol Exp Ther* 298:976
76. Yip-Schneider MT, Wiesenauer CA, Schmidt CM (2003) Inhibition of the phosphatidylinositol 3'-kinase signaling pathway increases the responsiveness of pancreatic carcinoma cells to sulindac. *J Gastrointest Surg* 7:354
77. Zhang X, Shu L, Hosio H, Murti KG, Houghton PJ (2002) Predominant nuclear localization of mammalian target of rapamycin in normal and malignant cells in culture. *J Biol Chem* 277:28127